

Package ‘ccrepe’

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Type Package

Title ccrepe_and_nc.score

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Imports infotheo (>= 1.1)

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Description The CCREPE (Compositionality Corrected by RENormalizaion and PERmutation) package is designed to assess the significance of general similarity measures in compositional datasets. In microbial abundance data, for example, the total abundances of all microbes sum to one; CCREPE is designed to take this constraint into account when assigning p-values to similarity measures between the microbes. The package has two functions:
ccrepe: Calculates similarity measures, p-values and q-values for relative abundances of bugs in one or two body sites using bootstrap and permutation matrices of the data.
nc.score: Calculates species-level co-variation and co-exclusion patterns based on an extension of the checkerboard score to ordinal data.

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VignetteBuilder knitr

Suggests knitr, BiocStyle, BiocGenerics, testthat

biocViews Statistics, Metagenomics, Bioinformatics, Software

R topics documented:

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ccrepe-package *A package for analysis of sparse compositional data. Allows calculation of the similarity measure nc-score and calculation of compositionality-corrected p-values for arbitrary similarity scores (including user-defined) applied to compositional data.*

Description

ccrepe was developed for use with microbial relative abundance data data, which is both sparse and compositional in nature.

Details

Package: ccrepe
 Type: Package
 Version: 1.0
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ccrepe *Calculates compositionality-corrected p-values and q-values for compositional data using an arbitrary distance metric.*

Description

ccrepe calculates compositionality-corrected p-values and q-values for compositional data by generating first a null distribution of the distance metric generated by permutation and renormalization of the data, and then by generating an alternative distribution of the distance metric by bootstrap resampling of the data. For greater detail, see References

The two distributions are compared using a pooled-variance Z-test to give a compositionality-corrected p-value. The p-values can be calculated for all appropriate (passing certain quality-control measures) pairwise comparisons, or for a subset of user-specified ones.

Q-values are additionally calculated using the Benjamin-Hochberg-Yekutieli procedure (see References)

Usage

```

ccrepe(
  x = NA,
  y = NA,
  sim.score = cor,
  sim.score.args = list(),
  min.subj = 20,
  iterations = 1000,
  subset.cols.x = NULL,
  subset.cols.y = NULL,
  errthresh = 1e-04,
  verbose = FALSE,
  iterations.gap = 100,
  distributions = NA,
  compare.within.x = TRUE,
  concurrent.output = NA,
  make.output.table = FALSE)

```

Arguments

- | | |
|----------------|---|
| x | First dataframe or matrix containing the relative abundances in cavity1 : columns are bugs, rows are samples. (Rows should therefore sum to a constant.)
The subjectIDs, if present, are assumed to be the row names and NOT the first column of data. |
| y | Second dataframe or matrix (optional) containing the relative abundances in cavity2: columns are bugs, rows are samples.
The subjectIDs, if present, are assumed to be the row names. If both x and y are specified, they will be merged by row names. If no row names are specified for either or both datasets, the default is to use the row numbers as subject IDs. |
| sim.score | A function defining a similarity measure, such as cor or nc.score. This similarity measure can be a pre-defined R function or user-defined. If the latter, certain properties should be satisfied as detailed below (also see examples). The default similarity measure is Spearman correlation.
A user-defined similarity measure should: <ol style="list-style-type: none"> 1.Be able to take either two inputs which are vectors or one input which is either a matrix or a dataframe 2.In the case of two inputs, return a single number 3.In the case of one input, return a matrix in which the (i,j)th entry is the similarity score for column i and column j in the original matrix 4.Resulting matrix (in the case of one input) must be symmetric 5.The inputs must be named x and y |
| sim.score.args | A list of arguments for the measurement function. For example: In the case of cor, the following would be acceptable: sim.score.args = list(method='spearman',use='complete.obs').
Note that this is the default behavior. |

<code>min.subj</code>	Minimum number of subjects that must be measured in a bug/feature/column in order to apply the similarity measure to that bug/feature/column. This is to ensure that there are sufficient subjects to perform a bootstrap (default: 20).
<code>iterations</code>	The number of iterations of bootstrap and permutation (default: 1000).
<code>subset.cols.x</code>	Subset of columns from x to work on. (Default: NULL - meaning: Use all columns of x. All the columns of x are used for normalization but calculations are performed only with the requested subset (by default, all columns).
<code>subset.cols.y</code>	Subset of columns from y to work on. (Default: NULL - meaning: Use all columns of y. If applicable (y present), all the columns of y are used for normalization, but calculations are performed only with the requested subset (by default, all columns).
<code>errthresh</code>	A numeric value representing the probability of getting all 0's in a given bootstrapped column for the first dataset. If a bug/feature/column has a number of zeros that makes the probability of obtaining all zeros when sampling with replacement > <code>errthresh</code> , that bug/feature/column will be excluded from the subsequent analysis. This is to ensure that the standard deviation of the bootstrap sample is non-zero. (default= 0.0001)
<code>verbose</code>	Logical: an indicator whether the user requested verbose output, which prints periodic progress of the algorithm through the dataset(s), as well as including more detailed output. (default:FALSE)
<code>iterations.gap</code>	If output is verbose - number of iterations after issue a status message (Default=100 - displayed only if <code>verbose=TRUE</code>).
<code>distributions</code>	Output Distribution file (default:NA).
<code>compare.within.x</code>	A boolean value indicating whether to do comparisons given by taking all subsets of size 2 from <code>subset.cols.x</code> or to do comparisons given by taking all possible combinations of <code>subset.cols.x</code> or <code>subset.cols.y</code> . If TRUE but <code>subset.cols.y=NA</code> , returns all comparisons involving any features in <code>subset.cols.x</code>
<code>concurrent.output</code>	Optional output file to which each comparison will be written as it is calculated.
<code>make.output.table</code>	A boolean value indicating whether to include table-formatted output.

Value

Returns a list containing the calculation results and the parameters used.

Default parameters shown:

<code>min.subj</code>	Description above
<code>errThresh</code>	Description same as <code>errthresh</code> above
<code>sim.score</code>	A matrix of the similarity scores for all the requested comparisons. The (i,j)th element of <code>sim.score</code> corresponds to the similarity score of column i (or the ith column of <code>subset.cols.1</code>) and column j (or the jth column of <code>subset.cols.1</code>) in one dataset, or to the similarity score of column i (or the ith column of <code>subset.cols.1</code>) in dataset x and column j (or the jth column of <code>subset.cols.2</code>) in dataset y in the case of two datasets.

p.values	A matrix of the p-values for all the requested comparisons. The (i,j)th element of p.values corresponds to the p-value of the (i,j)th element of sim.score.
q.values	A matrix of the Benjamini-Hochberg-Yekutieli FDR corrected p-values. The (i,j)th element of q.values corresponds to the q-value fo the (i,j)th element of sim.score.
z.stat	A matrix of the z-statistics for all the requested comparisons. The (i,j)th element corresponds to the z-statistic which gave rise to the (i,j)th p-value.
output.table	(Only if make.output.table=TRUE) A table where each row is one comparison. Each row contains the features being compared with their similarity scores, z-statistics, p-values and q-values

Additional parameters if verbose=TRUE:

iterations	Description Above
subset.cols.x	Description Above
subset.cols.y	Description Above
iterations.gap	Description Above
sim.score.parameters	Description Above
compare.within.x	Description Above
make.output.table	Description Above

Author(s)

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References

Emma Schwager and Colleagues. Detecting statistically significant associations between sparse and high dimensional compositioanl data. In Progress.

Benjamini and Yekutieli (2001). "The control of the false discovery rate in multiple testing under dependency." The Annals of Statistics. Vol. 19, No. 4. pp. 1165-1188.

Examples

```
data <- matrix(rlnorm(40,meanlog=0,sdlog=1),nrow=10)
data.rowsum <- apply(data,1,sum)
data.norm <- data/data.rowsum
testdata <- data.norm
dimnames(testdata) <- list(paste("Sample",seq(1,10)),paste("Feature",seq(1,4)))
ccrepe.results <-ccrepe (x=testdata, iterations=20, min.subj=10)
ccrepe.results.nc.score <- ccrepe(x=testdata,iterations=20,min.subj=10,sim.score=nc.score)
ccrepe.results
ccrepe.results.nc.score
```

```
ccrepeSampleTestFunction
```

ccrepeSampleTestFunction - Simple example of a test measurement function to be used with ccrepe

Description

This simple example of a test measurement function to be used with ccrepe used in the same fashion that cor would be used

Some properties of the function:

1. Be able to take either two inputs which are vectors or one input which is either a matrix or a data frame
3. In the case of one input, return a matrix in which the (i,j)th entry is the similarity score for column i and column j in the original matrix
4. Resulting matrix must be symmetric
5. The inputs must be named x and y

Usage

```
ccrepeSampleTestFunction(x, y = NA)
```

Arguments

x	x is a vector or a matrix
y	y is a vector. if y selected then x must be a vector too

Value

If x and y are vectors it returns a number: 0.5 If x is a matrix it returns a matrix of all 0.5

Author(s)

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nc.score

nc.score

Description

nc.score calculates species-level co-variation and co-exclusion patterns based on an extension of the checkerboard score to ordinal data.

It is an extension to Diamond's checkerboard score (See references below) to ordinal data and implements a framework for robust detection of species-level association patterns in metagenomic data. It performs quality control and transforms relative abundances to ordinal values and computes an association measure, the NC-score (n-dimensional checkerboard score), for each species pair.

Usage

```
nc.score(x = NA,
         y = NA,
         bins = NA,
         verbose = FALSE,
         min.abundance = 1e-04,
         min.samples = 0.1)
```

Arguments

x	A numeric vector, data frame, or matrix. The first entity to be processed. Columns are bugs, rows are samples.
y	Numeric vector. If selected, x must be a numeric vector as well.
bins	Either a single integer specifying the number of bins to use or a numeric vector specifying the cutoffs. If a single number is given, this is used as the number of bins in the discretize function of the package infotheo. If a vector is specified, the function findInterval is used to discretize the data according to the cutoffs given. The default behavior is to use the defaults for the discretize function.
verbose	Logical flag to request verbose output
min.abundance	A numeric value specifying the minimum abundance threshold: Assures selection of species with abundance \geq min.abundance in more than min.samples percent of samples.
min.samples	An fraction specifying the minimum samples threshold: Assures selection of species with abundance \geq min.abundance in more than min.samples percent of samples. Default=0.1

Value

Matrix or vector of normalized scores.

Author(s)

Craig Bielski<craig.bielski@gmail.com>

References

Emma Schwager and Colleagues. Detecting statistically significant associations between sparse and high dimensional composition data. In Progress.

Examples

```
data <- matrix(rlnorm(40,meanlog=0,sdlog=1),nrow=10)
data.rowsum <- apply(data,1,sum)
data.norm <- data/data.rowsum
testdata <- data.norm
dimnames(testdata) <- list(paste("Sample",seq(1,10)),paste("Feature",seq(1,4)))
```

```
nc.score.results <- nc.score( x=testdata )
nc.score.results.bins <- nc.score( x=testdata, bins=5 )
nc.score.results.bin.cutoffs <- nc.score( x=testdata, bins = c(.001,.25,.5) )
nc.score.results
nc.score.results.bins
nc.score.results.bin.cutoffs
```


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